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(21) International Application Number: PCT/US98/15937 (22) International Filing Date: 30 July 1998 (30.07.98) (30) Priority Data: 60/054,256 30 July 1997 (30.07.97) US (71) Applicant: CORNING INCORPORATED [US/US]; 1 Riverfront Plaza, Corning, NY 14831 (US). (72) Inventors: BOOKBINDER, Dana, C.; 2261 Davis Road, Corning, NY 14830 (US). HERSH, Leroy, S.; P.O. Box 449, Painted Post, NY 14870 (US). XIE, Xinying; 6 Overbrook Road, Painted Post, NY 14870 (US). (74) Agent: HERZFELD, Alexander, R.; Corning Incorporated, Patent Dept., SP FR 02-12, Corning, NY 14831 (US).		(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, HR, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZW, European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). Published <i>With international search report.</i>
(54) Title: OXIDIZED STYRENIC POLYMERS FOR DNA BINDING (57) Abstract A method of immobilizing aminated DNA to a substrate surface is provided for use in DNA hybridization assays. The surface can be widely employed in a variety of molded laboratory products including multiwell plates, cell culture dishes, and biological containers. The method comprises the steps of providing a polystyrene substrate having a surface content of between 0.3 and 4.0 % atomic percent oxygen, and attaching amine modified oligonucleotide to the surface of the substrate. A resultant product comprising a polystyrene substrate having a surface oxygen content of between 0.3 and 4.0 % atomic percent, and a plurality of amine modified oligonucleotides directly attached to the surface of the substrate, is also disclosed.		

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OXIDIZED STYRENIC POLYMERS FOR DNA BINDING

FIELD OF THE INVENTION

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The invention relates to laboratory ware for chemical and biological testing and experimentation and, more specifically, to a surface that will immobilize DNA.

BACKGROUND OF THE INVENTION

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Polymers have been widely utilized as vessels for cell culture, biological testing, medical research and other uses. Certain polymers such as styrenic polymers and more specifically, polystyrene, are optimal for use as substrates in specialty products for biological testing, and are typically employed. Polystyrene, for example, is cheap,
15 optically clear, and may be processed at low temperatures.

Direct immobilization of DNA to a substrate surface has many applications to a variety of assay systems in molecular biology, such as immuno-PCR or detection of nucleic acids with specifically labeled probes, i.e. target capture hybridization. Target capture hybridization relies on immobilizing capture oligonucleotide molecules (usually
20 single strand DNA or RNA fragments that may be isolated from a natural source, or produced by standard molecular biological techniques, having from about two to forty nucleotides) on a support surface and then hybridizing labeled complementary target DNA strands to the immobilized molecules.

An aminated nucleic acid molecule is said to be "immobilized" to a solid support if it is either adsorbed to the support, or covalently bonded thereto, with sufficient strength that it cannot be removed from the support by washing with water or an aqueous buffer. A certain release of capture oligonucleotides invariably occurs with systems employing adsorption as the method of attachment. Therefore, covalent bonding of the oligonucleotide is preferred.

One method of immobilizing aminated DNA by covalent bonding involves attaching dialdehyde starch (DAS) or N-oxysuccinimide esters (NOS) to a polymer surface. In surfaces having a DAS coating, the NH_2 that is attached to the aminated capture oligonucleotide covalently links to an aldehyde group from the DAS, thereby immobilizing the oligonucleotide molecule to the polystyrene surface via the coating. A method of attaching DAS to the surface of a cell culture plate is disclosed in United States Patents 5,563,215 and 5,281,660.

With a NOS coating, DNA with primary amines added synthetically or by in-vitro manipulation can be directly coupled to the NOS surface. When the primary amines react with NOS groups at a slightly alkaline pH, the ester undergoes nucleophilic substitution as the amine attacks the carbonyl group and displaces the N-oxysuccinimide group. The specificity of this reaction creates the immobilization effect.

Another technique for immobilizing DNA to a substrate surface is by covalently grafting secondary amines to a polystyrene surface. In this technique, the oligonucleotide is first phosphorylated at the 5' end with polynucleotide kinase and ATP. The phospholated DNA is then coupled to the secondary amines on the surface using carbodiimide for 5 hours at 50°. The DNA is thereby immobilized by a phosphoramidate bond.

A problem exists with surface treatment involving coatings such as DAS or NOS, and with attaching secondary amines. With time, or when subjected to various environmental stresses, the surface becomes unstable and the immobilization characteristics of the attached coating are diminished. Further, the coupling efficiency of binding aminated DNA to such treated plates is quite low, less than 0.1%. Also, the surface treatment of the polymer substrate adds an additional step to the manufacturing process and adds cost to the final product.

The present invention provides a method of using a product that is made of relatively pure polystyrene that covalently binds aminated DNA. The product works as well as substrates that have been treated with DAS or NOS, but without the cost of adding a coating. The present invention can also be subjected to a variety of environmental stresses without losing any of its immobilizing characteristics. Additionally, since the product of present invention is reactive toward amine groups in general, it is capable of binding peptides and low molecular weight antigens for immunoassays, as well as antibodies, cell attachment factors and other bioreactive molecules.

SUMMARY OF THE INVENTION

The present invention provides a method of immobilizing aminated DNA to a substrate surface. The surface can be widely employed in a variety of molded laboratory products including multiwell plates, cell culture dishes, and biological containers. The method comprises the steps of providing a polystyrene substrate having a surface content of between 0.3 and 4.0 atomic percent oxygen, and attaching amine modified oligonucleotide to the surface of the substrate. A resultant product comprises a polystyrene substrate having a surface oxygen content of between 0.3 and 4.0 atomic percent, and a plurality of amine modified oligonucleotides directly attached to the surface of the substrate.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1 is a graph of the effect that various surfaces have on binding aminated DNA.

FIG. 2 is a graph comparing the binding effects of NOS coated polystyrene with the binding effects of an untreated polystyrene strip.

FIG. 3 is a graph of the effect that surface oxygen content of polystyrene has on ability to bind aminated DNA while using binding solutions of varying pH.

FIG. 4 is a graph of the effects of pH on ability to bind aminated DNA.

FIG. 5 is a graph of the effects of humidity on ability to bind aminated DNA.

FIG. 6 is a graph comparing the immobilization of both unmodified and amine modified cDNA on oxidized polystyrene and polystyrene coated with NOS.

FIG. 7 is a graph comparing the immobilization characteristics of both optically clear and opaque black polystyrene with solutions having varying concentrations of aminated DNA.

FIG. 8 is a graph comparing aminated cDNA binding on optically clear and opaque black reactive polystyrene.

DETAILED DESCRIPTION OF THE INVENTION

The present invention results from an accidental discovery by the inventors. A study was conducted by the inventors to investigate the mechanisms involved in binding aminated DNA to various surfaces. Aminated DNA refers to a DNA oligimer that has been modified by addition of a primary amine attachment on a terminal end. The inventors used polystyrene multiwell strips coated with both NOS and DAS and the results were to be compared to an uncoated polystyrene strip of wells which was to serve as the control. "Multiwell strips" or "strip of wells" consist of a number of wells (typically 8), of the type which comprise the wells in multiwell plates, which are attached to each other to form a linear array. U.S. patent 3,649,464 shows a multiwell plate and U.S. patent 5,084,246 shows a multiwell strip. To the amazement of the inventors, the uncoated, untreated control strips bound the DNA as well as the treated strips. The inventors then began work to derive an explanation for this seemingly anomalous result. As a first step, they noted that all other studies that used untreated polystyrene controls were performed using multiwell plates instead of well strips, and no significant binding had been detected on the plates.

Having made this observation, the inventors then embarked on a study to compare various substrate surfaces to determine their ability to bind aminated DNA. The substrates they selected were: (1) plain, untreated polystyrene plates and strips, sold under the tradename "MEDIUM BIND" available from Corning Costar Corporation, (2) polystyrene plates and strips that had been subjected to 3 mRads of gamma radiation,

sold under the tradename "HIGH BIND" available from Corning Costar Corporation, (3) polystyrene plates and strips coated with NOS, and (4) tissue culture plates made of polystyrene that had been subjected to Corona discharge. It should be noted that the tradenames "HIGH BIND" and "MEDIUM BIND" refer to the product's propensity for binding protein in ELISA assays; it does not refer to ability to bind aminated DNA.

The procedure used for colorimetric detection of biotinylated DNA as used to obtain the data on binding was as follows:

1. To the plate or strip, 100 ul/well of amine modified probe oligonucleotide in oligo binding buffer (50 mM Na_2PO_4 , pH 8.5, 1mM EDTA) was added. The combination was either incubated overnight at 4° C or for 1 hour at 37° C.

2. Uncoupled oligonucleotide was removed by washing the plate three times with PBS.

3. The unreacted active groups were blocked by adding 200 ul/well of 3% BSA in oligo binding buffer. The plate or strip was then incubated for 30 minutes at 37° C and then decanted.

4. 100 ul/well of hybridization solution containing the target nucleic acid was then added. The plate or strip was then incubated for 1 hour at 49° C.

5. The wells were washed with preheated 2XSSC (3.0M NaCl; 0.3M sodium citrate, pH 7.0) 0.1% SDS twice and soaked for 5 minutes at 49° C.

6. 100 ul/well of blocking solution containing streptavidin-peroxidase conjugate diluted 1:1000 was added. The plate or strip was incubated for 30 min. at 37° C.

7. The wells were washed three times with PBS.

8. 100 ul/well of fresh substrate solution was added. OD readings were taken at 405 nm at 10, 20, and 30 minutes. The OD readings were uncorrected for background absorbance.

In the experiment that yielded the results shown in FIG. 1, aminated DNA was applied to plates at a concentration of 1.0 pmol/well and incubated for 1 hour at 37° C.

The results were recorded 20 minutes after adding fresh substrate solution. The readings that were taken are averages of readings from four different wells. Noise is a reading taken without DNA probes attached, signal is a reading measured with DNA probes. The background noise readings were obtained by following all the above listed

preparation steps, but without adding any probe DNA. By subtracting this noise level from the signal obtained when probe DNA was added, all non-specific binding signal can be eliminated and a true measure of the binding effectiveness can be obtained.

As can be observed from FIG. 1, the untreated polystyrene strips (Med St.)
5 bound the aminated DNA more effectively than the untreated plate (Med Pl.) or any of the treated surfaces.

As a further illustration, an experiment was performed the results of which are displayed in FIG. 2. Aminated DNA was applied to NOS coated polystyrene plates and untreated polystyrene strips at various concentrations in oligo binding buffer (for NOS
10 coated plates, pH 8.5, 1M NaCl, 50mM phosphate, 1mM EDTA; for untreated strips, 0.1M phosphate, pH 11.0) and incubated for 1 hour at 37° C. The values were obtained 20 minutes after adding substrate solution by averaging four wells and reading the OD at 405nm. As the results show, the untreated polystyrene strip exhibits more signal, especially at lower concentrations, and therefore better binding characteristics than the
15 NOS coated surface under these conditions.

The inventors next sought to determine what was different about the untreated strips when compared to the untreated plates from the same polystyrene. It was observed that the multiwell strips used in the experiment were produced by a molding process that was slightly different than the molding process used in the formation of the
20 multiwell plate. For one, in molding the strips, no lubricant was used in the molds. Conversely, the molding of a plate requires lubricant, such as zinc stearate, in the mold cavity for proper ejection of the ware. Second, the mold for the strip uses more air venting than molds for the plates. With this in mind, the inventors began to look closely at the surface chemistry of both the untreated strips and plates, and more specifically,
25 the surface oxygen content of the polystyrene.

The process of making the well strips, it was discovered, creates a product that has a surface oxygen content of approximately 0.6%, while plates molded with lubricant and less venting have approximately four times less surface oxygen. Subsequent testing has revealed that an optimal level of surface oxygen exists for creating a substrate that
30 effectively binds or immobilizes aminated DNA.

FIG. 3 shows the relationship between surface oxygen and ability to bind aminated DNA. The aminated DNA was applied to polystyrene surfaces at a concentration of 1.0 pmol/well in binding solution having four different pH levels, and incubated for 1 hour at 37 °C. The ability to bind aminated DNA is measured by the amount of signal received back from a labeled oligonucleotide effectively bound to the surface. These results, which represent averages of readings taken from four different wells, were obtained 20 minutes after adding substrate solution and taken at 405 nm. Table 1 displays the testing results that are plotted in FIG. 3.

Table 1

Substrate	Atomic % Oxygen	Signal pH 3	Signal pH 6	Signal pH 8.5	Signal pH 11
Abraded PS Strip	0.11	N/A	N/A	0.32	N/A
Untreated PS Plate	0.15	0.04	0.17	0.30	0.73
Untreated PS Strip	0.62	0.27	2.04	2.17	2.19
Treated PS Plate (Treated w/ 1.5 mRads of Gamma Radiation)	0.81	1.83	2.10	1.99	2.31
High Treated Plate (Treated w/ 3.0 mRads of Gamma Radiation)	1.34	1.61	2.11	1.39	2.38
High Treated Strip (Treated w/ 3.0 mRads of Gamma Radiation)	3.04	1.18	1.58	0.44	2.14
Tissue Culture Treated Plate	10.0	0.11	0.00	0.01	0.01

As seen from FIG. 3, conditions for binding aminated DNA are optimized in the untreated polystyrene strips at approximately 0.3-2.0 atomic percent, varying slightly depending on the pH of the binding solution. Of the four pH levels surveyed, using an oligo binding buffer having a pH of 11 provided the best binding results at each of the test points.

When the polystyrene strip was abraded so that the surface that contacted the mold was scraped away, both oxygen content and binding ability were decreased, thus substantiating the importance of oxygen on the surface. When polystyrene plates were irradiated with 1.5 mRads of gamma sterilization, the atomic percentage of oxygen and

the binding ability of the plates increased significantly. However, when polystyrene is subjected to higher doses of gamma radiation, i.e. 3.0 mRads, the surface oxygen content increases, but binding ability tails off. This data suggest an optimum level of surface oxygen content for binding aminated DNA.

5 It should be noted that although the optimal surface oxygen atomic percentage is approximately between 0.3 and 4.0, binding ability is still enhanced to some degree for surface oxygen atomic percentages between 4.0 and 8.0 when using binding solution having a pH of 3, 6, or 11 as shown in FIG. 3. Additionally, some advantageous binding occurs with polystyrene having a surface oxygen atomic percentage between 0.2 and
10 0.3. Polystyrene having a binding signal that is below 0.5 has essentially lost its utility as a product for use in assays requiring the immobilization of aminated DNA.

 Aside from molding polystyrene products by the process used in making the well strips, surface oxygen content can be increased by several methods known to those skilled in the art. One example is by treatment with appropriate amounts of gamma
15 radiation. Other known methods for affecting surface oxygen content include UV exposure, ozone treatment, and corona discharge. The surface oxygen content can be varied based on intensity and duration of these treatments.

 FIG. 4 illustrates the effect that pH plays on binding ability of uncoated polystyrene plates having an optimal atomic percentage of surface oxygen. In the
20 experiment that led to the results displayed in FIG. 4, the aminated DNA was applied to plates at a concentration of 1.0 pmol/well and incubated for 1 hour at 37° C with different pH buffers. The results were obtained 20 minutes after adding fresh substrate solution and readings were taken by OD at 405nm. The plotted values are averages of results from four different wells. The results indicate that, unlike with surfaces coated
25 with DAS or NOS, aminated DNA will bind to the surface at a variety of pH levels.

 FIG. 5 illustrates the effect that humidity, over a period of time, has on aminated DNA binding. An untreated polystyrene strip and a strip and plate coated with NOS were stored at 85% humidity at 25° C for up to 4 weeks before the assay. The aminated DNA was applied to the plates at a concentration of 1.0 pmol/well and incubated for 1
30 hour at 37° C. The results were obtained 20 minutes after adding fresh substrate solution and readings were taken by OD at 405nm. The plotted values are averages of

results from four different wells. As can be seen from FIG. 5, the untreated polystyrene strips bound the aminated DNA better, at each testing point, than the strips coated with NOS.

Data generated indicates further that the linker arm attaching the primary amine to the DNA molecule is essential for DNA binding to untreated polystyrene having surface oxygen atomic percent between 0.3 and 4.0. As used in the experiments described herein, carbon chain molecules of lengths between 6 and 12 carbon atoms were used as linker arms. It has been observed that without the linker arm, less than 25% of signal is obtained when compared to binding results with the linker arm. It has been further observed that the length of the linker arm does not significantly affect DNA binding. This data suggests that the immobilization of aminated DNA on the polystyrene having the optimum surface oxygen content, very likely occurs by chemical or physical bonding between the NH_2 and the polystyrene surface. It also eliminates the possibility of a strong contribution of hydrophobic binding between the non-polar carbon-carbon chain in the linker arm and the non-polar polystyrene surface.

The results from the pH experiments, which are displayed graphically in FIG. 4, also shed some light on the binding mechanism between the polystyrene and aminated DNA. The results showed that there were two maxima peaks, at pH 6 and pH 11. This suggests that there might be two different mechanisms involved in the binding of aminated DNA to the surface of polystyrene having a surface oxygen content of between 0.3 and 4.0 atomic percent. The maxima at pH 6 could indicate either physical or chemical bonding -- ionic bonding between the NH_3^+ group on the aminated DNA and carboxylate groups on the oxidized polystyrene surface or chemical reaction between the NH_2 group on the aminated DNA and carbonyl groups on the oxidized polystyrene. The pH maxima at pH 11 may be due to chemical bonding between the amine groups on the aminated DNA and the carboxyl groups on the oxidized polystyrene surface.

Nevertheless, knowledge about the mechanisms behind the binding of aminated DNA to the surface of polystyrene having the requisite surface oxygen content is not required to practice the invention.

Further, and as displayed graphically in FIG. 6, polystyrene having a surface oxygen content of 0.62 atomic percent, bound amine modified cDNA significantly better

than NOS coated polystyrene or unmodified cDNA when using a binding solution of pH 11. This shows that even aminated DNA strands having greater than 100 nucleotides (cDNA) may be effectively immobilized on polystyrene having an optimum surface oxygen content. Further, even the unmodified cDNA molecules bound to the oxidized polystyrene better than the modified cDNA molecules bound to the NOS coated surface. This perhaps suggests that in the cDNA molecules, amine groups from the numerous individual nucleotides participate in the immobilization.

It should be understood that the binding effects of a polystyrene surface having an optimum level of surface oxygen, as described, is not limited to aminated DNA. The surface is reactive toward amine groups in general. This implies that it is possible to derivitize such reactive polystyrene with various kinds of pendent groups such as carboxyl, amine, biotin, etc. As a result, larger bioreactive molecules such as antigens, antibodies and cell attachment factors may be covalently attached and thereby immobilized to the polystyrene surface. In fact, the reactive polystyrene will react with cell attachment peptides to provide an effective cell growth surface for an endothelial cell line. This approach can also extend to include polyethylene oxide tethered peptides.

One application of the present invention is to pattern polystyrene with alternating cell binding and cell non-binding, or alternating regions of DNA binding and DNA non-binding, in forming an assay array. The individual binding regions may be the subject of independent testing. The alternating regions can be made, for example, by providing polystyrene having non-binding characteristics (surface oxygen content preferably outside of the 0.3 - 8.0 atomic percent range), and treating only selected portions of the surface to gamma radiation sufficient to create binding characteristics (surface oxygen content preferably between 0.3 - 4.0 atomic percent). The regions of the surface that are designated as non-binding may be shielded from the radiation by, for example, a patterned mask.

It is also believed that the unique immobilization characteristics shown in relation to surface oxygen content in the range of 0.3-8.0 atomic percent extend to other styrenic polymers such as styrene acrylonitrile copolymer (SAN), ABS (acrylonitrile-butadiene-styrene), butadiene-styrene copolymers (HIPS and MIPS, K-resin and Kraton(R)), polyparamethyl styrene, Styrene-acrylic copolymer, poly alpha methyl styrene, and

polymer blends containing styrenic based polymers. The preliminary testing done to date on these polymers has confirmed this belief.

5 It should also be noted that optically clear polystyrene having a surface oxygen content of 0.62 atomic percent (MEDIUM BIND), was compared with black opaque styrene-butadiene copolymer (MIPS) having a surface oxygen content of 0.62 atomic percent, for oligonucleotide binding effects. As can be deduced from the immobilization data graphically displayed in FIG. 7 and FIG. 8, no significant difference exists between opaque black and optically clear polystyrene in its ability to bind aminated DNA or cDNA. What this indicates is that the pigment used in forming the opaque MIPS do not
10 have a detrimental effect on surface binding. This has substantial relevance for the potential use of opaque polystyrenes in chemiluminescent or fluorescent immobilization assays that rely on opaque substrates to limit background noise.

Although the invention has been described in detail for the purpose of illustration, it is understood that such detail is solely for that purpose and variations can
15 be made therein by those skilled in the art, such as modifying the surface oxygen content of other polymers, without departing from the spirit and scope of the invention which is defined by the following claims.

WE CLAIM:

1. A method of immobilizing DNA to a surface comprising:
 - a) providing a styrenic polymer substrate having a surface oxygen content of
5 between 0.2 and 8.0 atomic percent, and
 - b) attaching a DNA to the surface of the substrate.
2. The method of claim 1 wherein said DNA is an amine modified oligonucleotide.
- 10 3. The method of claim 1 wherein said DNA is an amine modified cDNA.
4. The method of claim 1 wherein said DNA is an unmodified cDNA.
5. The method of claim 1 wherein said styrenic polymer substrate has a surface
15 oxygen content of between 0.3 and 4.0 atomic percent.
6. The method of claim 1 wherein said styrenic polymer substrate has a surface
oxygen content of between 0.3 and 2.0 atomic percent.
- 20 7. A product for use in chemical and biological analysis comprising:
a styrenic polymer substrate having a surface oxygen content of between 0.2 and
8.0 atomic percent; and
a plurality of DNA molecules directly attached to the surface of said substrate.
- 25 8. The method of claim 7 wherein said DNA molecules are amine modified
oligonucleotide.
9. The method of claim 7 wherein said DNA molecules are amine modified cDNA.
- 30 10. The method of claim 7 wherein said DNA molecules are unmodified cDNA.

11. The product of claim 7 wherein said styrenic polymer substrate has a surface oxygen content of between 0.3 and 4.0 atomic percent.
12. The product of claim 7 wherein said styrenic polymer substrate has a surface oxygen content of between 0.3 and 2.0 atomic percent.
13. The product of claim 7 wherein said substrate is a multiwell plate.
14. The product of claim 7 wherein said substrate is a strip of wells.
15. A method of immobilizing an aminated biological molecule to a surface comprising:
- a) providing a styrenic polymer substrate having a surface oxygen content of between 0.2 and 8.0 atomic percent, and
 - b) attaching amine bearing biological molecule to the surface of the substrate.
16. The method of claim 15 wherein said styrenic polymer substrate has a surface oxygen content of between 0.3 and 4.0 atomic percent.
17. The method of claim 15 wherein said styrenic polymer substrate has a surface oxygen content of between 0.3 and 2.0 atomic percent.
18. The method of claim 15 wherein said biological molecule is a peptide.
19. The method of claim 15 wherein said biological molecule is an antigen.
20. The method of claim 15 wherein said biological molecule is an antibody.
21. The method of claim 15 wherein said biological molecule is a cell attachment factor.

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OF INTERNATIONAL PROCESSING

31. The product of claim 29 wherein said binding regions have a surface oxygen content of between 0.3 and 2.0 atomic percent.

5 32. The product of claim 29 wherein said biological molecules are amine modified oligonucleotides.

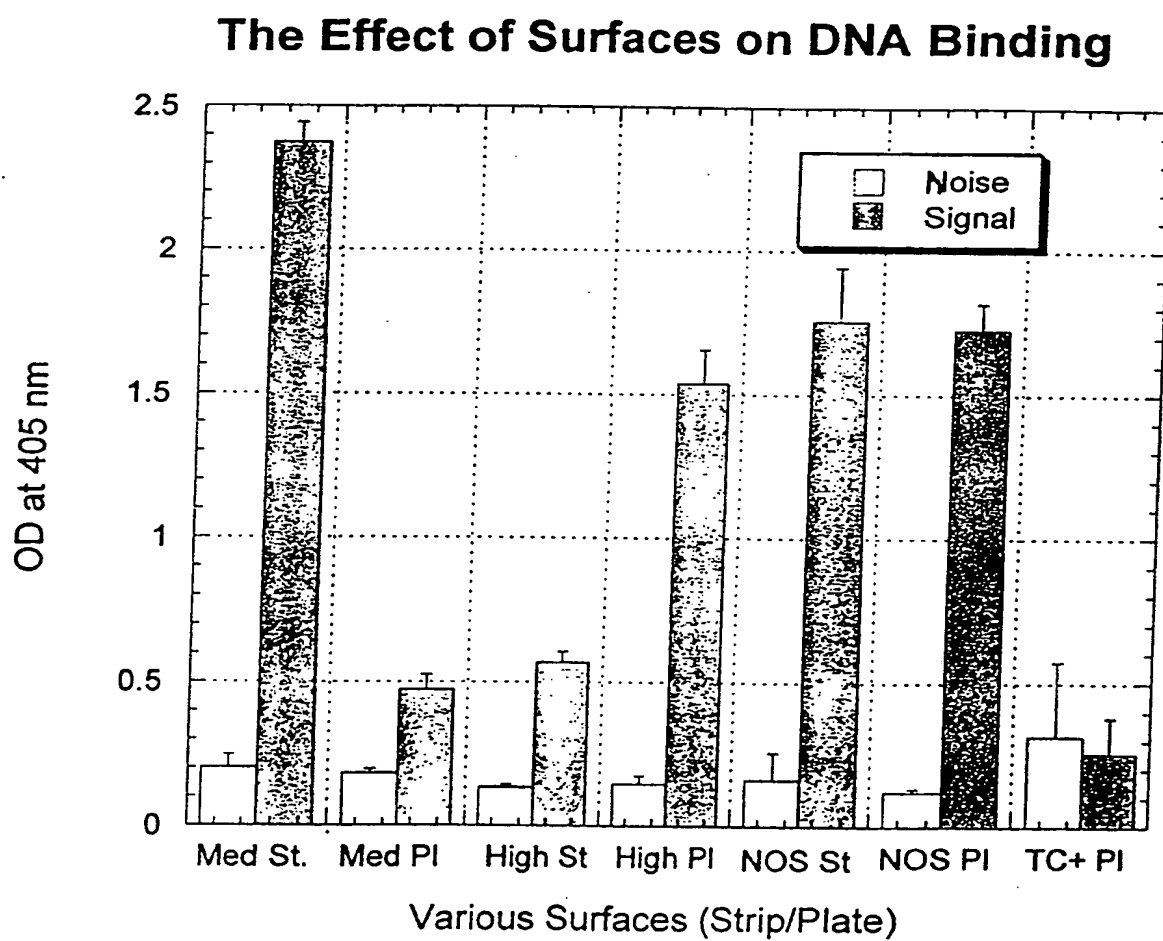
33. The product of claim 29 wherein said biological molecules are peptides.

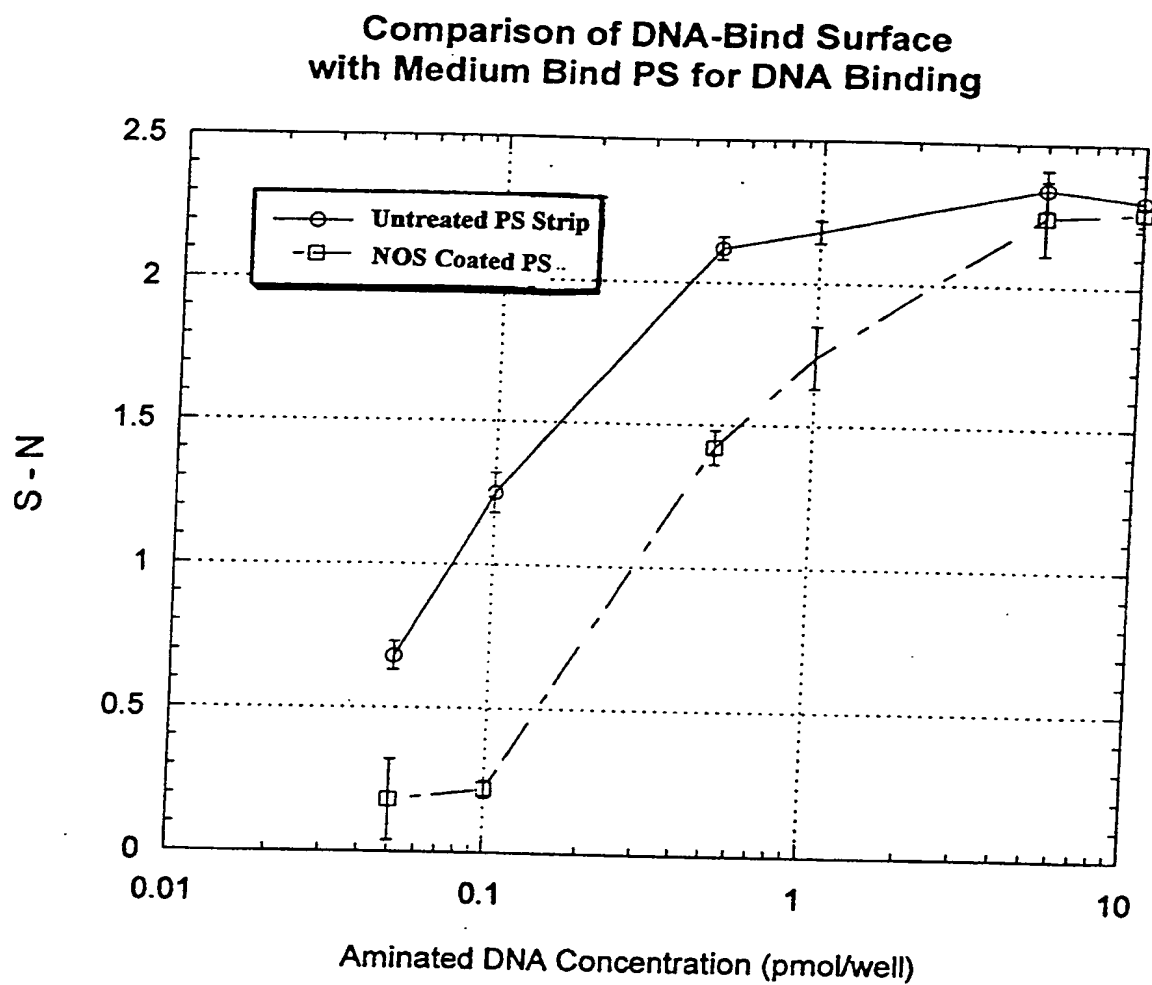
10 34. The product of claim 29 wherein said biological molecules are antigens.

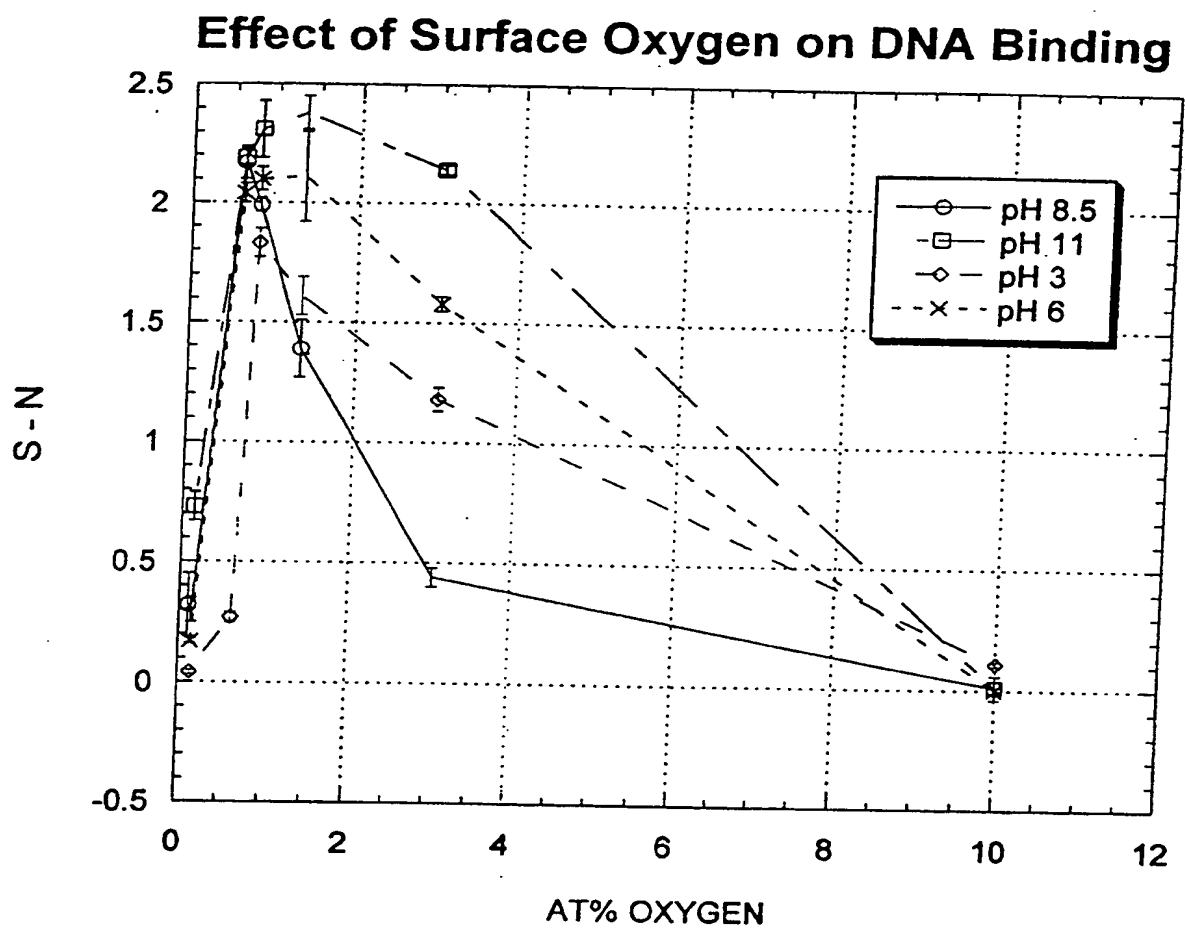
35. The product of claim 29 wherein said biological molecules are antibodies.

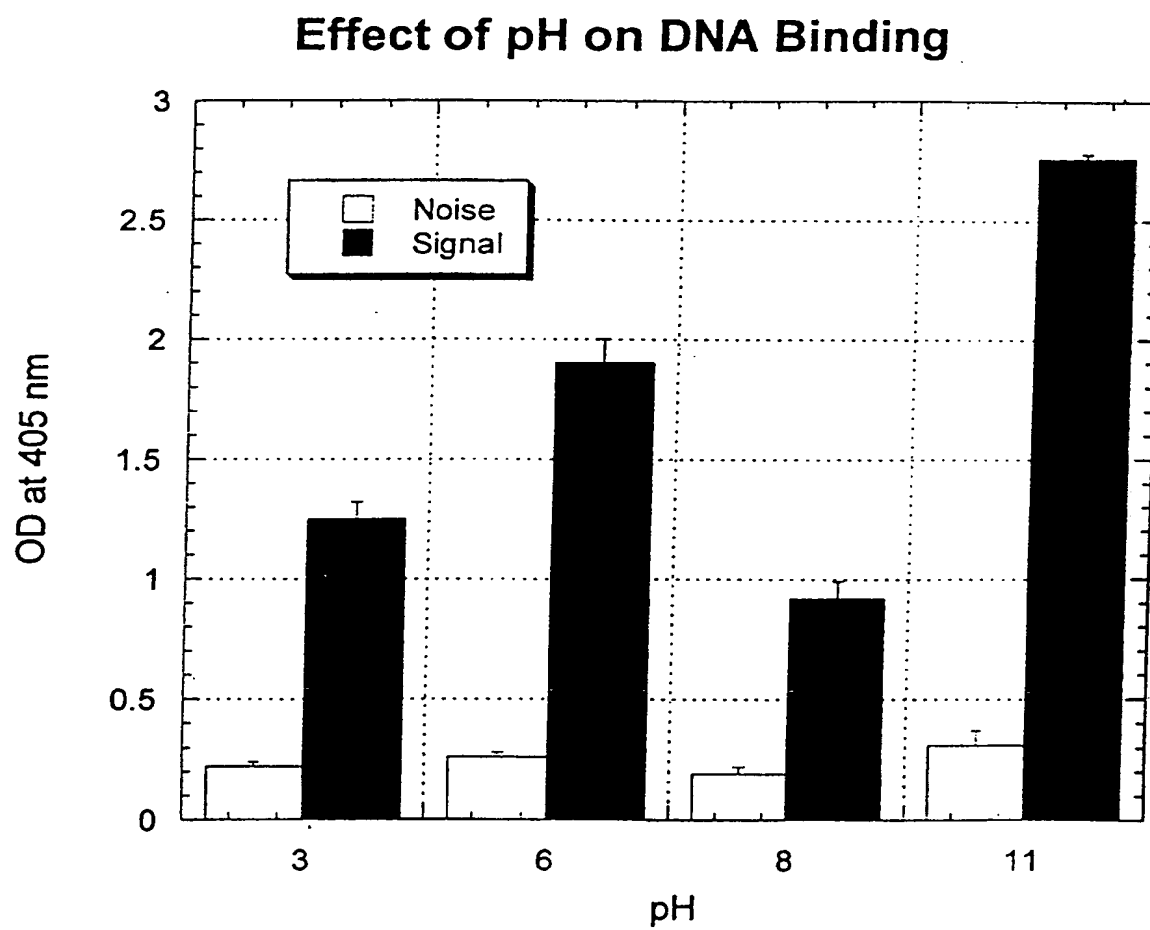
15 36. The product of claim 29 wherein said biological molecules are cell attachment factors.

37. A method of making a substrate for use in chemical or biological testing comprising:
a) providing a polymeric styrene substrate having a surface, and
b) subjecting said surface to between 0.1 and 3.0 mRads of gamma
20 sterilization.

**FIG. 1**

**FIG. 2**

**FIG. 3**

**FIG. 4**

The Effect of Humidity on DNA Binding

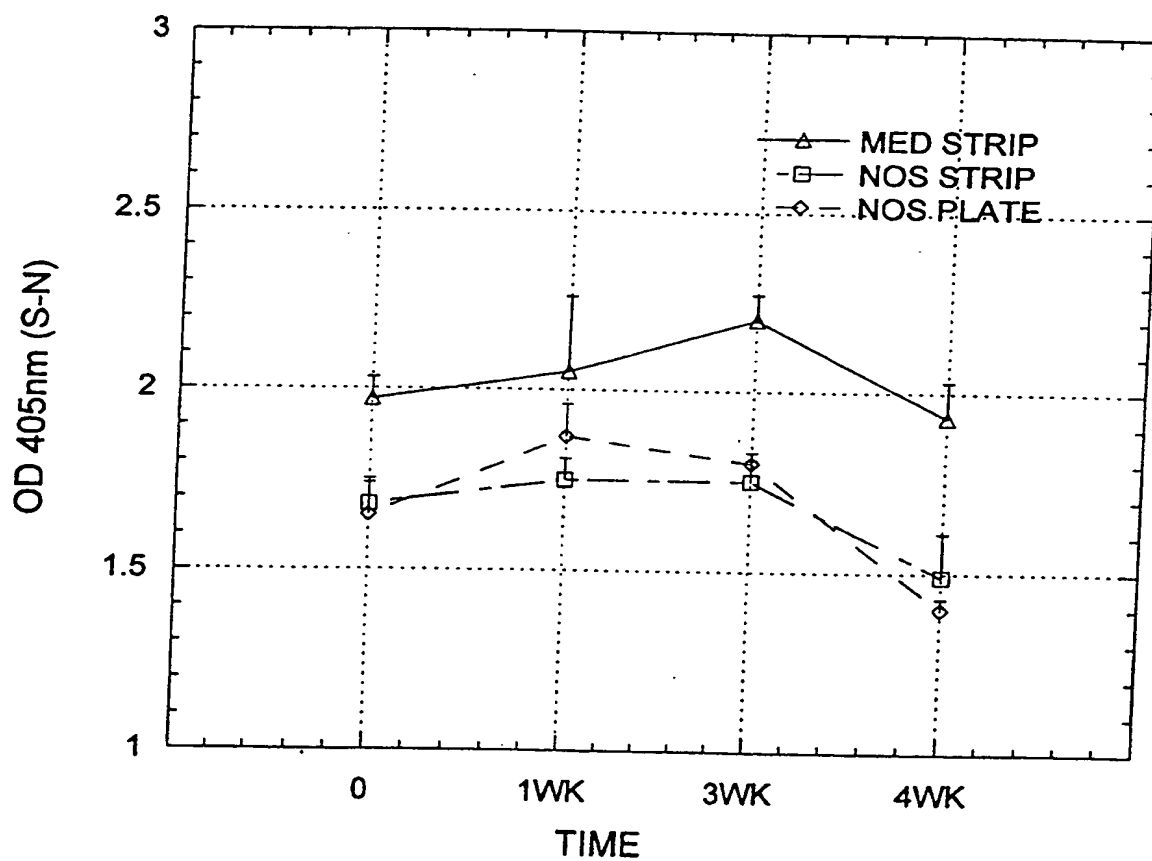
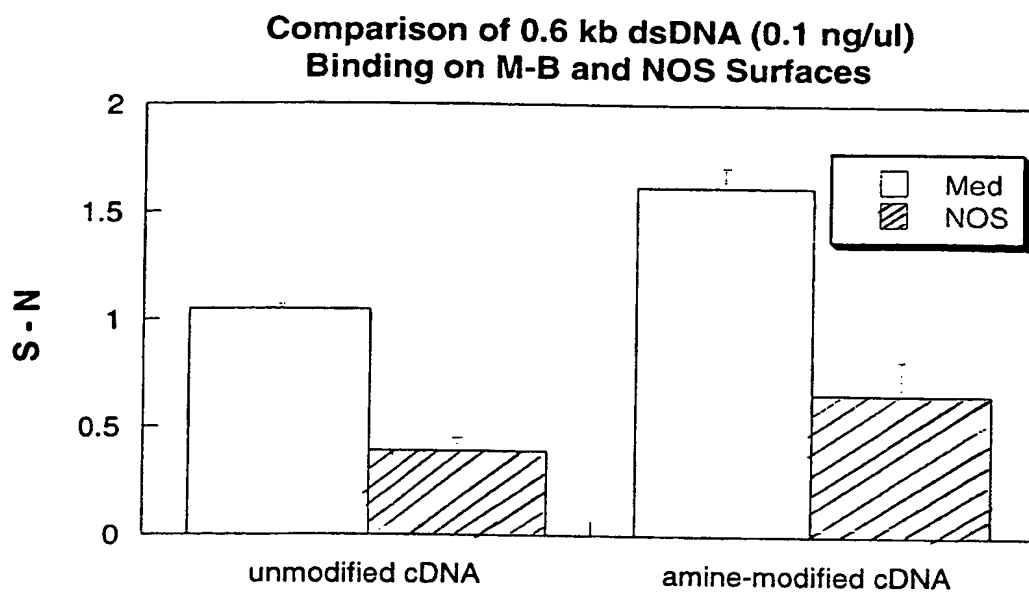
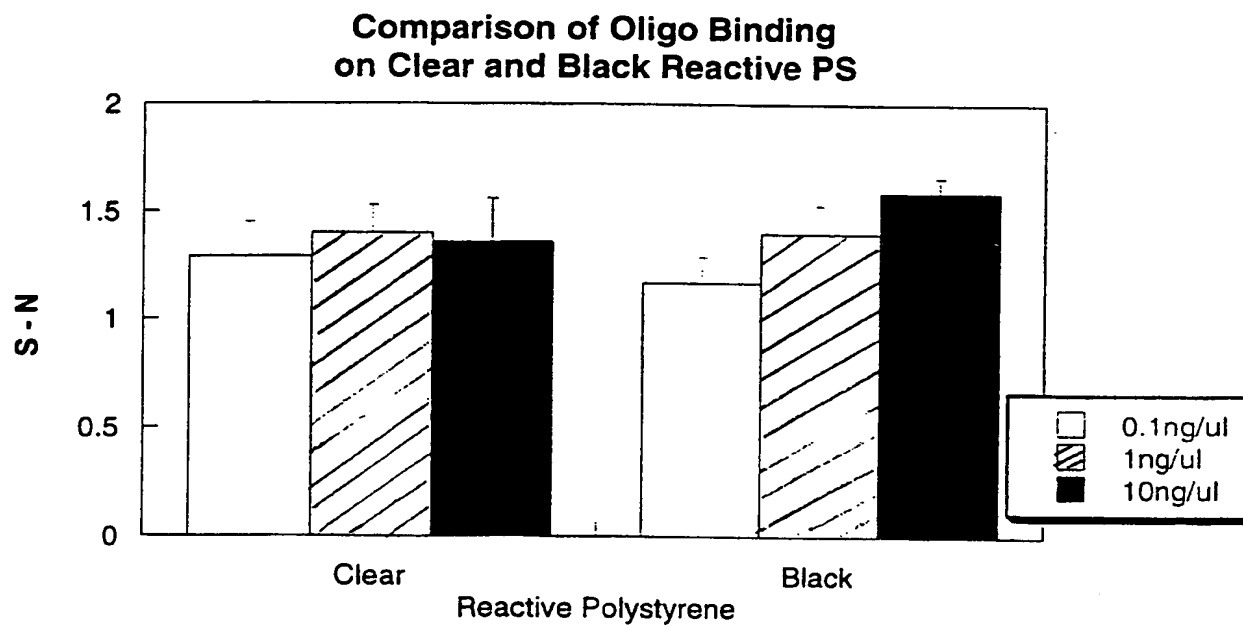
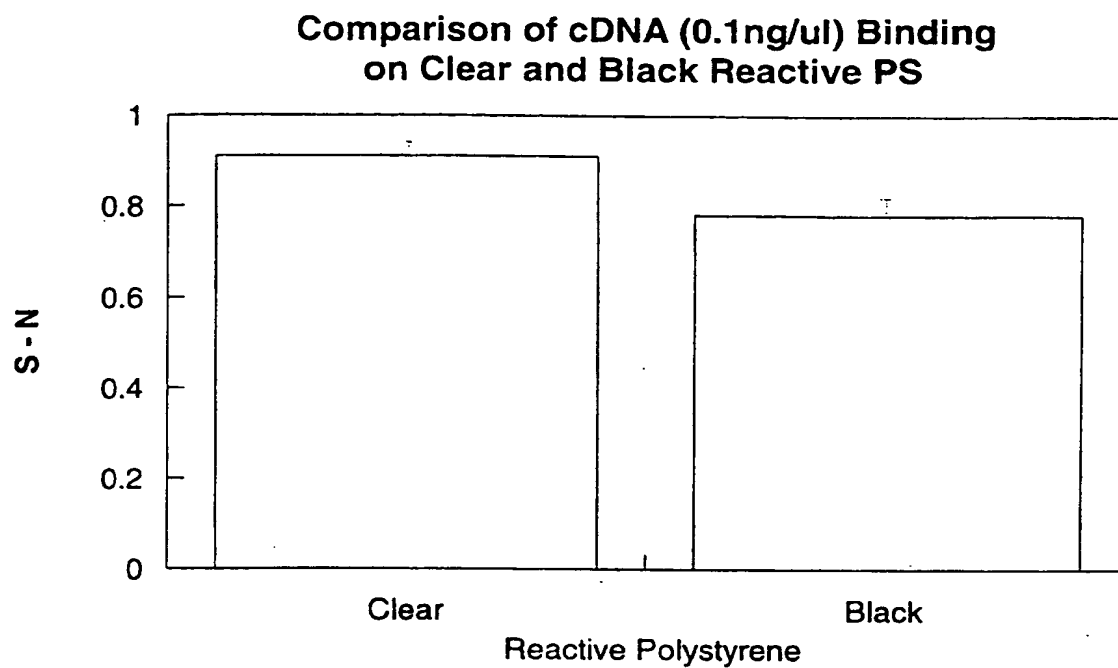


FIG. 5

**FIG. 6****FIG. 7**

7 / 7

**FIG. 8**

INTERNATIONAL SEARCH REPORT

 International application No.
PCT/US98/15937

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) :C07H 21/00; C08G 63/91; G01N 35/539, 544, 545

US CL :525/54.2; 536/23.1; 436/528, 531, 532, 535

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 525/54.2; 536/23.1; 436/528, 531, 532, 535

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
NONEElectronic data base consulted during the international search (name of data base and, where practicable, search terms used)
NONE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 4,789,628 A (NAYAK) 06 December 1988, see entire document.	1-14
Y	US 4,994,373 A (STAVRIANOPOULOS et al) 19 February 1991, see entire document.	1-14
Y	US 5,089,387 A (TSAY et al) 18 February 1992, see entire document.	1-14
Y	UA 5,262,297 A (SUTTON) 16 November 1993, see entire document.	1-14
Y	US 5,414,075 A (SWAN et al) 09 May 1995, see entire document.	1-14

☐ Further documents are listed in the continuation of Box C.
 ☐ See patent family annex.

* Special categories of cited documents:	*T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
*A document defining the general state of the art which is not considered to be of particular relevance	*X document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
*E earlier document published on or after the international filing date	*Y document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
*L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*A document member of the same patent family
*O document referring to an oral disclosure, use, exhibition or other means	
*P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

08 OCTOBER 1998

Date of mailing of the international search report

30 OCT 1998

 Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
Box PCT
Washington, D.C. 20231

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Authorized officer

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INTERNATIONAL SEARCH REPORTInternational application No.
PCT/US98/15937**Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)**

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

1-14

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US98/15937

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

I) Group I, claims 1-14, drawn to a method for immobilizing DNA to a surface and a product of that method, classified in Class 525, subclass 54.2, and Class 536, subclass 23.1.

II) Group II, claims 15-28, drawn to a method of immobilizing an aminated biological molecule to a surface and a product of that method, classified in Class 436, subclasses 528, 531, 532 and 535, Class 525, subclass 54.11 and Class 530, subclass 815.

III) Group III, claims 29-36, drawn to a styrenic polymer substrate having an array of alternating binding and non-binding regions, classified in Class 525, subclass 335.

IV) Group IV, claim 37, drawn to a method of making a substrate for chemical or biological testing by subjection of the surface with radiation, classified in Class 522, subclasses 150, 157, 158 and 160.

The inventions listed as Groups I, II, III and IV do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: The listed inventions lack unity of invention under PCT Rules 13.1 to 13.3 because there is no special technical feature which links the diverse modified surfaces of either of the groups. The surface of Group I is modified with a different species and by a different method than that of Group II. Each of the methods are drawn to different process steps.